

# **Yield, transpiration efficiency, and water use variations and their relationships in the sorghum reference collection**

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**Abstract.** Sorghum is well adapted to water limited conditions, but the traits responsible for this enhanced adaptation under drought conditions remain unclear. In this study, yield, transpiration efficiency and water extraction were assessed in 149 germplasm entries from the sorghum reference set (plus three control cultivars) using a lysimetric system under terminal water stress and fully irrigated conditions outdoors. A ten-fold range for grain yield and harvest index (HI), two-fold range for transpiration efficiency (TE) and a 1.25 fold variation for water extraction were observed under terminal water stress conditions. Transpiration efficiency and water extraction under water stress related poorly to that under fully irrigated conditions, reflecting a large genotype-by-water treatment interaction. Under drought stress, total water extraction varied by about 3 L plant<sup>-1</sup> among germplasm. Entries from the Durra race had highest water extraction capacity, whereas Caudatum-Bicolor and Caudatum-Durra intermediate races had poor water extraction. Durra, Caudatum and Caudatum-Guinea races had highest TE, whereas the Guinea race had the lowest. Although yield was closely related to HI, at any level of HI there were substantial yield differences that remained unexplained, and these residual yield variations were closely related to transpiration efficiency ( $R^2 = 0.60$ ). Similarly, substantial yield variations that were still not explained by HI or TE were closely related to the total water extracted under water stress ( $R^2 = 0.35$ ). A multilinear regression analysis confirmed these results and showed the importance of water extraction during grain filling. Therefore, next to HI, the yield differences under terminal drought in

sorghum were driven by TE, and then next by water extraction. The large genetic variation for TE and water extraction offer great breeding opportunities and, in particular, highlight the Durra race as a critical source of variation.

**Additional keywords:** Roots, germplasm reference set, water uptake profile, pre-anthesis water use.

## Introduction

Water deficit is the most important abiotic stress and significantly limits crop production globally, particularly in the Semi-Arid Tropics (SAT). There are different “patterns” of water stresses depending on the timing, the intensity, and the duration of drought stress (Serraj *et al.* 2005). In the SAT, where the length of the cropping period is limited, sorghum often faces a terminal drought, caused by the cessation of rain toward the end of the rainy season. This is particularly the case for post-rainy (*rabi*) sorghum in India, which is sown at the end of the rainy season to take advantage of the moisture accumulated in the soil profile. Successful crops under terminal drought are those having increased water availability and accessibility during grain filling (Vadez *et al.* 2007a). Possible options for increasing water availability post-anthesis are to: (i) manage the soil moisture profile in a way that leaves water available for grain filling, including strategies to minimize water use before anthesis (Kholova *et al.* 2010a, 2010b) or strategies to enhance transpiration efficiency; (ii) develop a deeper and/or more profuse rooting system to access extra water from the soil profile.

Having higher transpiration efficiency (TE, in g biomass kg<sup>-1</sup> water transpired) could contribute to a slower rate of soil moisture depletion. Genotypic differences for TE have been reported in sorghum under well watered conditions (Hammer *et al.* 1997; Xin *et al.* 2009). Few studies have looked at TE under both fully irrigated and water stress conditions (Donatelli *et al.* 1992; Balota *et al.* 2008), with only a limited range of germplasm being assessed. Also, except Balota *et al.* (2008), TE has been measured over relatively limited periods of time. So it is important to assess genetic variation for TE

over an entire crop cycle and to determine whether there are large genotype-by-water regime interactions for TE. We used this approach to assess a large and diverse set of germplasm lines from the sorghum reference set (Ramu *et al.* manuscript in preparation).

Rooting traits have been reported as potentially important for drought adaptation in sorghum (Bhan *et al.* 1973, Mayaki *et al.* 1976, Blum *et al.* 1977; Jordan *et al.* 1979) based on studies involving a limited number of genotypes. In one study, the roots of a drought tolerant sorghum line grew at least 40 cm deeper than a drought sensitive one (Salih *et al.* 1999). Yet, root measurements are time-consuming and prone to error (Vadez *et al.*, 2007a), and the range of genotypic variation for the capacity to extract water from a soil profile remains unknown. This is critical information to gather since recent simulation and experimental work in wheat shows that each millimeter of water extraction during the grain filling period contributes to an additional 55 kg ha<sup>-1</sup> grain yield (Manschadi *et al.* 2006) and 59 kg ha<sup>-1</sup> grain yield (Kirkegaard *et al.* 2007), respectively. It was also shown that the total water extraction did not differ between the tolerant and the sensitive wheat genotype, but the tolerant line used less water before anthesis and more after anthesis than the sensitive line (Manschadi *et al.* 2006). Here, similar hypotheses are developed in sorghum to assess differences in the total water extraction and the proportion of water being used during the post-anthesis period to service grain filling (Hammer 2006).

Passioura's equation (1977) ( $Y = WU \times TE \times HI$ , with Y, WU, TE, and HI standing for yield, water used, transpiration efficiency and harvest index) has been widely used to guide the search for traits contributing to drought adaptation. However, since there was no method to evaluate all components on the same plants with an equal degree of precision, the use of that equation was generally limited to only single components, regardless of the relative importance of other components. For example, the past twenty years of drought research in groundnut has focused on water use efficiency (Hubick *et al.* 1986; Wright *et al.* 1994; Nageswara Rao *et al.* 2001; Udaykumar *et al.* 1998 Krishnamurthy *et al.* 2007), often relying on surrogates to estimate trait value. Similarly, rooting traits have been used as surrogates for water extraction (the WU component) (reviewed in Vadez *et al.* 2007a). Whether high TE relates to low water use (Blum *et al.* 2005) or not (Peng and Krieg 1992) is still a matter of debate. Also, it is possible that one

of the components of the equation may have, under specific conditions, a greater bearing on yield, thereby obscuring the true contribution of the other components to yield. Here, a method is used (Vadez *et al.* 2008; Ratnakumar *et al.* 2009) to precisely assess all components of Passioura's equation on same plant and test their relationships using a large set of germplasm.

Sorghum is among the most adapted crops to dryland farming. Yet there is considerable genetic diversity available for adaptation to water deficit (Crasta *et al.* 1999; Harris *et al.* 2007). Recently, a mini core collection of 242 accessions of sorghum germplasm lines representing global diversity in core and entire collections has been developed using data on 21 phenotypic traits (Upadhyaya *et al.* 2009). More recently, a reference set collection based on data from 41 SSR markers consisting of 384 entries has been developed (Ramu *et al.* manuscript in preparation). We assess variation in the traits described above in a sub-set of the reference set of sorghum which was chosen to limit variation in their time to flowering.

The overall objective of the present study was to assess variation in the sorghum reference collection for traits related to plant water use and hypothesized to be closely related to crop adaptation under terminal drought. We specifically assessed: (i) the genotypic differences in water extraction and the pattern of water use before and after anthesis; (ii) the genotypic variation in transpiration efficiency; (iii) the range of water regime-by-genotype interaction for these traits; (iv) the contribution of these traits to grain yield under terminal drought, and (v) possible association between specific sorghum races and values of the traits assessed.

## Materials and methods

### Soil filling and growth conditions of the lysimeters

Plants were grown in lysimeters, i.e. PVC tubes of 25 cm diameter and 2.0 m length, filled with Alfisol in outdoor conditions, with possibility to cover them with a shelter in case of rain. A PVC end plate was placed on top of four screws at the bottom of the cylinders, 3cm from the very bottom, to prevent the soil from seeping through. The

endplate did not fit the cylinder tightly and allowed water drainage, although drainage was prevented when lysimeter weighing started (see below). Tubes were filled with Alfisol collected from the ICRISAT farm and sieved to particles smaller than 1 cm. This allowed the bulk density of the soil profile to be set at approximately  $1.35 \text{ g cm}^{-3}$ , a standard value for Alfisols. Cylinders were filled with soil in three increments of 40 kg of dry soil. After addition of each 40 kg increment, the soil level in several cylinders was checked to ensure they were similar in all tubes. Then each 40 kg of soil added was watered. A previous assessment of the water needed to fill the profile before drainage determined that the soil water holding capacity of the Alfisol was approximately 20%. Therefore, 8 L of water were added to each 40 kg increment of soil. After adding/watering 40 kg of soil three times, an additional 15 kg of dry soil was added to each cylinder and watered with 3 L. At that stage, the cylinders were almost filled to the desired level, i.e. approximately 5 cm from the top. A top-up using dry soil was done to ensure that all cylinders were filled to the same level. This top-up varied between about 1-2 kg, i.e. less than 1-2% variation across cylinders. Hence, all the cylinders had a similar bulk density close to  $1.35 \text{ g cm}^{-3}$ . All cylinders at field capacity weighed between 163 and 165 kg.

The soil in the lysimeters had been fertilized with DAP and muriate of potash, both at a rate of  $200 \text{ mg kg}^{-1}$  soil. It was also complemented with sieved and sterilized farm manure at a rate of 2:50 to prevent micro-nutrient deficiency. Before growing the sorghum crop, the lysimeters were used for a crop of finger millet and foxtail millet, planted sequentially. The foxtail millet crop had received a urea top-dressing of  $3 \text{ g plant}^{-1}$ . At the end of this crop, only the main root stock from the plants was removed from the top layer of soil by softening the top soil with water and pulling. The soil was then tilled superficially with sickles and limited Alfisol top-up was added so that the surface level was approximately 5 cm below the lysimeter brim. This created a soil profile that was undisturbed from previous cropping, except for minimum tillage of the surface. The lysimeters were then watered to field capacity, based on their expected weight, and the sorghum crop was planted on a full profile. The crop was top dressed with  $3 \text{ g urea plant}^{-1}$  at 4 weeks after sowing.

## 150 Space arrangement of the lysimeters and weighing

151 The top of the cylinders was equipped with a metal collar and rings that allowed them to  
 152 be lifted. Weighing of the cylinders was done by lifting the cylinders with a block-  
 153 chained pulley, and an S-type load cell (Mettler-Toledo, Geneva, Switzerland) was  
 154 inserted between the rings of the cylinder and the pulley. The scale (200 kg capacity)  
 155 allowed repeated measurements with an accuracy of 20 g on each weighing. The  
 156 lysimeters were separated from one another by a distance of approximately 5 cm. Thus  
 157 the sorghum crop was planted at a density of approximately 11 plants m<sup>-2</sup>, a plant  
 158 population similar to typical field plantings at ICRISAT (row-to-row distance of 60 cm  
 159 and plant-to-plant spacing of 15 cm). This allowed us to accurately assess the water  
 160 extraction pattern of a crop cultivated in conditions similar to the field. The tubes were  
 161 arranged in four trenches of 2 m depth and 1.75 m width. Each trench was separated by a  
 162 20 cm concrete wall. Possible border effects were expected on the south side of the  
 163 trenches (these were oriented east-west) and those effects were curbed by bordering the  
 164 trench with two rows of plants on the south side of the trenches.

165

## 166 Treatments used and trait assessed

167 Four seeds were planted in each cylinder on 20 October 2008 during the *Rabi* sorghum  
 168 season. Plants were thinned to two seedlings per cylinder at 14 days after sowing (DAS)  
 169 and then to one plant per cylinder at 21 DAS. Disturbance to remaining plant was  
 170 avoided by clipping the thinned plant below the collar. All plants were fully irrigated  
 171 until 28 DAS. This involved cylinders receiving 500 mL twice a week for the first two  
 172 weeks after sowing, and then on alternate days until 28 DAS. At 28 DAS, the cylinders  
 173 were covered with a 2-cm layer of low density polyethylene beads to prevent soil  
 174 evaporation. Preliminary testing indicated that the beads prevented more than 90% of the  
 175 soil evaporation, so that differences in mass primarily reflected plant transpiration (data  
 176 not shown). Biomass increase between weighing was negligible compared to plant water  
 177 use. Weighing of the cylinders was done at 30 DAS for the first time and then  
 178 subsequently every two weeks. This gave a total of five weights until harvest for the DS  
 179 plants and six weights for the WW plants. The first weighing at 30 DAS gave the field

capacity weight of each cylinder. The cylinders were distributed in four trenches and the weighing of one trench per day was done. The same sequence of weighing was used for each trench so that the time intervals between weighing were the same in all cylinders.

To keep the WW plants sufficiently wet for optimum growth and to avoid water drainage after irrigation, the WW plants were watered when the cylinder weight, at the time of weighing, had fallen below 2L from the weight at field capacity. This prevented drainage at the bottom. The watering was done every week. The week that plants were not weighted, the water addition of the previous week was used for watering the WW plants. The DS treatment received no water from 28 DAS until maturity, except for 2L that were added to all cylinders at 73 DAS (beginning of grain filling).

Flowering time (d) was recorded on a plant basis. Transpiration was calculated at approximately 2-weekly intervals between 31 DAS (the time at which weighing started) and maturity. Daily transpiration values were calculated for each plant by dividing the transpiration for each time interval between weighing by the number of days in each interval. Pre-anthesis transpiration was the sum of the daily transpiration values until anthesis, plus water used in the first 28 days after sowing, which was estimated to be 1.5L for all genotypes. This was based on dry biomass estimates of 15 g at 28 DAS and on the assumption of a TE of  $10 \text{ g kg}^{-1}$  water transpired at such an early stage (our unpublished observations). The post-anthesis water use was the sum of the daily transpiration values from anthesis until maturity. Harvest was done over a period of 4 days. Leaf, stem (including sheath) and panicle masses were recorded after drying for 3 days in a forced-air oven set at  $70^\circ\text{C}$ . Panicles were then subsequently threshed to determine grain yield. The harvest index was calculated as the ratio of grain yield divided by the total aboveground biomass (the aggregated mass of stems, leaves, and panicles). Transpiration efficiency (TE) was calculated as the ratio of the total aboveground biomass divided by the sum of transpiration values between 30 DAS and maturity. The initial biomass at the time of initiating the transpiration measurements was not taken into account, assuming that biomass differences between genotypes at that stage were negligible. This would have led to a slight over-estimation of TE.

## 210 Plant material

211 The flowering time of 384 lines belonging to the sorghum reference set had been  
 212 determined under field conditions in 2008-09 (Upadhayaya, pers. comm.). Based on these  
 213 data, 149 reference set entries and three control cultivars, IS 2205, IS 18758, and IS  
 214 33844, varying in flowering time between 70 and 85 days after sowing, were selected. IS  
 215 2205 is a Durra-Bicolor landrace resistant to shoot fly and stem borer. IS 18758 is a  
 216 Guinea-Caudatum landrace, released as E 35-1 in Burkina Faso in 1983 and as Gambella  
 217 1107 in Burundi in 1990. IS 33844 is a Durra landrace released in India as Parbhani Moti  
 218 in 2002. The 149 reference set lines represented 30 out of 44 countries in the entire  
 219 reference set. Race-wise composition was Caudatum (31), Durra (18), Bicolor (17  
 220 accessions), Guinea (14), Kafir (6), Guinea-Caudatum (24), Caudatum-Bicolor (14),  
 221 Durra-Caudatum (13), Durra-Bicolor (3), Kafir-Bicolor (1), and Kafir-Caudatum (1). An  
 222 accession each of *aethiopicum* and *virgatum*, two accessions of *drummondii*, and three  
 223 of *verticilliflorum* were also part of the 149 reference set material.

224 In addition to the DS and WW sets of plants used above, a third set of plants was sown  
 225 at the same time in an area adjacent to the trenches. Plants were grown in 25 cm pots  
 226 filled with 11 kg of the same Alfisol. Previous experiments in sorghum using these pots  
 227 showed no signs of growth restriction due to pot size up to anthesis. The same planting  
 228 procedures were used and plants were kept well-watered until harvest. This set was  
 229 harvested at flowering and its purpose was to evaluate leaf area and tillering  
 230 characteristics of the different genotypes at that stage.

231

## 232 Data analysis

233 The experiment design was an Alpha lattice with 19 blocks of 8 entries within each  
 234 block. There were three replications and two water regimes (WW and DS). The Residual  
 235 Maximum Likelihood (ReML) method of Genstat was used to obtain the unbiased  
 236 estimate of the variance components and the best linear unbiased predictions (BLUPs) for  
 237 the different parameters measured within each treatment, considering genotypes as  
 238 random and replications as fixed effects. The significance of the genetic variability  
 239 among accessions within treatment was assessed from the standard error of the estimate



of genetic variance  $\sigma^2_g$ . Analysis was also performed to assess the effect of genotype (G), water treatment (T) and genotype-by-water treatment (GxT) interaction for the different traits measured. In this case, G and GxT were considered as random effects whereas treatment and replication were considered as fixed effects. The significance of genetic variability across treatments or of the genotype-by-treatment interaction effect was assessed in a manner similar to the above. The significance of the fixed effect of the treatment was assessed using the Wald statistic that asymptotically follows a  $\chi^2$  distribution.

For the multi-linear regression analysis, a multi-linear model was used in the software STATA (Stata Corp. College Station, Tx, USA), where yield was taken as an additive function of HI, TE, total water extraction, water extracted in the post-anthesis period, water extracted in the 45-59 DAS and 59-78 DAS period, days to flowering, and a constant. The same multi-linear model was used to assess the residual yield variations not explained by HI (see below), therefore excluding HI from the list of explanatory variables.

## Results

### *Yield and biomass components*

Grain yield varied significantly between genotypes under DS conditions, ranging from 0.3 to 36.6 g plant<sup>-1</sup> (Fig. 1a). Overall, the mean yield of 20.6 g plant<sup>-1</sup> under DS conditions was about 50% of the yield mean under WW conditions (42.0), indicating that the stress imposed was neither too severe nor too mild (Table 1). Under WW conditions, grain yield varied from 2.1 to 82.8 g plant<sup>-1</sup>. Grain yield under WW and DS conditions were poorly related ( $R^2 = 0.10$ ), which also reflected the large genotype-by-treatment (GxT) interaction for grain yield (Table 1).

Harvest index (HI) also varied significantly between genotypes under DS conditions, ranging from 0.05 to 0.52 (Fig. 1b), except for two genotypes that did not produce any grains. The overall mean HI of 0.27 under DS was only slightly smaller than the mean HI under WW conditions (0.33). The HI also varied considerably under WW conditions,

ranging from 0.21 to 0.53, except for five genotypes that had a poor HI lower than 0.15. Contrary to the grain yield data, the HI under DS conditions was better related to the HI under WW conditions ( $R^2 = 0.33$ ) (Fig. 1b), although HI also displayed a significant genotype-by-water treatment interaction (Table 1). The total plant biomass varied largely between entries. Under DS conditions, there was a two-fold difference between the minimum and the maximum value, whereas under WW conditions these differences were about four-fold. This reflects in part genotypic differences in plant size and tillering, which became larger under WW conditions. Since the genotypes were randomized in the different replications, it is also a possibility that dwarf germplasm may have suffered from shading from tall germplasm in the WW conditions. This possibility is, however, quite unlikely under DS conditions, where the range of total biomass was smaller than under WW conditions, and also where total biomass differences also reflected large differences in grain yield.

#### *Total water extraction*

Total water extracted under DS conditions varied significantly ( $P < 0.001$ ) among the 149 entries, ranging from 10,600 to 15,200 g plant<sup>-1</sup>. Noticeably, a low CV of only 6% was obtained for the total water extraction in the lysimetric system. Under fully irrigated conditions, the water extracted by the plants also varied significantly, ranging from 10,500 to 42,300 g plant<sup>-1</sup>. Besides an expected treatment effect, the total water extracted showed a large GxT interaction effect (Table 2), whereas the genotypic effect was non-significant. In fact, the water extracted under WW and DS conditions showed a poor relationship ( $R^2 = 0.08$ ). Total water uptake under DS conditions was assessed for each individual race. The Durra race had the highest total water uptake (14,120 g plant<sup>-1</sup>, n=20) (Fig. 2). The Durra-Caudatum race had, on average, the lowest total water uptake (13,570 g plant<sup>-1</sup>, n=12), followed by the Caudatum-Bicolor accessions (13,800 g plant<sup>-1</sup>, n=14).

The first two water use measurements for the 31-45 DAS and 45-59 DAS time intervals were similar in WW and DS plants (Fig. 3), although there was a significant, but minor, treatment effect on the water extraction in the 45-59 DAS interval (Table 2). Indeed, the water used under DS in the 45-59 DAS period was above 70% of that under WW

conditions, except for 18 lines where water used was 50-70% of that under WW. This indicated that for the 29 days following the last irrigation in the DS treatment, DS plants extracted similar amounts of water to WW plants. Water uptake in the 59-78 DAS interval showed a large treatment (T) effect on water extraction, and large and significant G and GxT effects, the latter being more important than the G effect. Water uptake in the 78-94 DAS interval also showed large T and GxT effects, and no significant G effect. By 59 DAS, 125 out of 152 entries had flowered and all the others flowered by 65 DAS.

Summarising, the large variation in water extraction capacity under DS conditions, with a tendency to have higher water extraction in Durra race than in Durra-Caudatum race, resulted from specific adaptation of genotypes to the stress conditions, and the temporal pattern of water use indicated that stress occurred after flowering for most lines.

#### *Relationships between water extracted before and after anthesis*

The pre-anthesis water use varied by 9 L plant<sup>-1</sup> among genotypes (5 to 14 L plant<sup>-1</sup> range). These differences were, in part, explained by the flowering time ( $R^2 = 0.70$ , data not shown) although large variations in pre-anthesis water use per day, which removes the differences due to flowering time, remained (101 to 205 g water per day). Pre-anthesis water use was also significantly correlated to the leaf area at anthesis ( $R^2 = 0.18$ , data not shown). Pre-anthesis water use under DS was also predominantly determined by genetic effects (Table 2). The post-anthesis water used ranged from about 2 to 10 L plant<sup>-1</sup> among genotypes. Post-anthesis water use under DS was correlated to flowering time ( $R^2 = 0.73$ ) but not to the post-anthesis water use of WW plants. Pre- and post-anthesis water use showed a close negative correlation ( $R^2 = 0.83$ ) (Fig. 4). Post-anthesis water use was also predominantly determined by genotype-by-treatment interaction effects, whereas the G effects were not significant (Table 2). Post-anthesis water use was also negatively correlated to the leaf area at anthesis ( $R^2 = 0.17$ ). These data indicate that despite flowering time determining about two thirds of the variation in pre- and post-anthesis water use, there was still a large range of variation in pre- and post-anthesis water use at any level of flowering time.

### 329 *Transpiration efficiency*

330 Transpiration efficiency varied largely among entries, ranging from 3.21 to 6.09 g kg<sup>-1</sup>  
 331 water transpired under DS conditions. The coefficient of variation was only 13.6%.  
 332 Under fully irrigated conditions, TE also varied significantly, ranging from 2.95 and 5.59  
 333 g kg<sup>-1</sup> (Table 1). The grand mean of 4.30 g kg<sup>-1</sup> was lower than under DS conditions (4.82  
 334 g kg<sup>-1</sup>). TE under DS and WW conditions were correlated but the correlation coefficient  
 335 was weak ( $R^2 = 0.13$ , data not shown). Besides a strong treatment effect on TE, G and  
 336 GxT interaction effects were both significant although the magnitude of the Ge effects  
 337 was slightly higher. Transpiration efficiency was assessed for each individual race under  
 338 DS conditions. The Guinea race exhibited the lowest mean TE values (4.29g kg<sup>-1</sup>, n=13),  
 339 followed by the Kafir (4.58g kg<sup>-1</sup>, n=6), whereas the Guinea-Caudatum, Durra and  
 340 Caudatum races had the highest mean TE values (5.09, 5.05 and 4.98 g kg<sup>-1</sup>, n = 25, 20  
 341 and 32, respectively) (Fig. 5). In summary, TE was mostly driven by genotypic effects  
 342 rather than by GxT interactions, and high TE variants were identified, especially in the  
 343 Guinea-Caudatum, Durra and Caudatum races.

344

### 345 *Relationships between water extraction, TE, HI, and yield*

346 Regression analyses were conducted between grain yield and WU, TE, and HI. The  
 347 relationship between grain yield and water used was significant under fully irrigated  
 348 conditions only ( $R^2 = 0.33$ ), but not under DS conditions (data not shown). Similarly,  
 349 grain yield was significantly related to TE under WW conditions ( $R^2 = 0.35$ ) and,  
 350 although the relationship was significant under DS conditions, the correlation coefficient  
 351 was weak ( $R^2 = 0.07$ ) (data not shown).

352 Therefore, individually, neither the total water used nor TE had any substantial bearing  
 353 on yield under DS conditions. This was because the relationship between yield and HI  
 354 was highly significant, and more so under DS conditions ( $R^2 = 0.88$ ) than under WW  
 355 conditions ( $R^2 = 0.53$ ) (Fig. 6a). However, for any given HI level, Figure 6a indicates  
 356 clearly that substantial variation in yield remained unexplained by HI, especially at HI  
 357 levels above 0.30. These residual yield variations unexplained by HI were calculated by  
 358 subtracting the yield predicted by the regression equation (Fig. 6a) from the observed

yields, following Vadez et al (2007b). These residuals showed a highly significant correlation with TE ( $R^2 = 0.60$ ) (Fig. 6b) and the total water extracted ( $R^2 = 0.40$ ; data not shown). Similarly, for any given TE level, yield variation remained unexplained by HI and TE. These residual yields were calculated in a similar way, using the regression equation of Fig 6b. These residual yields, unexplained by either the HI or TE, were closely related to the total water used ( $R^2 = 0.43$  or  $0.35$ , excluding or including an outlier data point on the left of the regression curve) (Fig. 6c). Other regressions were tested between these residuals and the pre- and post-anthesis water use, and the pre- and post-anthesis water used normalized by the flowering time, but no significant relationships were found (data not shown).

Similar results were observed from a multi-linear regression, where an additive model was used (Table 3), among several others that were tested. The model explained 98% of the phenotypic variation in yield. In that analysis, HI was the explanatory variable accounting for the largest component of yield, followed by TE and water used. In this model, the post-anthesis water use had a significant negative effect on yield, while the water extracted in the 59-78 DAS period, corresponding to the flowering stage of most genotypes, had a significant positive effect on yield. Finally, the time to flowering had a significant negative effect on grain yield (Table 3). The same approach was used to explain the residual yield variations not explained by HI. The best model explained 79% of the variation in the residuals and parameters having the most influence were TE and the total water use, with TE having a probability coefficient about twice that of total water use (Table 4). In that model, the water extracted in the 45-59 DAS and 59-78 DAS intervals were both significantly and positively related to the residual yield variations.

In summary, besides a strong HI effect on yield under DS conditions, the large yield variations remaining unexplained by HI were mostly driven by TE, and then next by total water extraction differences.

## Discussion

Our data showed a two-fold range of variation for TE and almost 20% variation for water extraction ( $3L\ plant^{-1}$ , equivalent to 30 mm on a field basis) under water stress in this

selection of lines from the sorghum reference collection. Water extraction and TE measurements in the lysimeters were reliable, exhibiting very low CVs (6% and 13.6%, respectively). After removing the proportion of yield differences explained by HI, there was still a substantial yield variation unexplained, and this was highly significantly related to TE. Water extraction only ranked third when accounting for yield variations unexplained by either HI or TE. High genetic variation for TE and water extraction offer substantial breeding opportunities, with high TE and water extraction variants in the Durra race being a critical source of key terminal drought adaptation traits for sorghum. Importantly, high TE and water extraction are not necessarily mutually exclusive, as evidenced by variants of the Durra race.

#### Large genotypic differences in TE

Transpiration efficiency was increased by about 10% under DS compared with WW conditions and since the GxT interactions were rather small, differences were driven by genotypic effects. This contrasted with previous results reporting a higher TE under fully irrigated conditions (Donatelli *et al.* 1992). Also the range of variation reported here under both water regimes was almost two-fold higher than in previous work reporting 20% (Donatelli *et al.* 1992) and 25% (Hammer *et al.* 1997) differences in TE. The values reported here were slightly below the range found by Balota *et al.* (2008) (5.04 – 7.55 g kg<sup>-1</sup> water) which is the only reported study where TE was measured over the entire crop cycle. The values here were also slightly below that in Hammer *et al.* (1997) (6.0 – 7.7 g kg<sup>-1</sup>). Many of these differences could simply be related to differences in VPD under different conditions. Also roots were not included in our TE assessment (inclusion of roots could increase absolute TE values by about 30-40%). There was no significant relationship between TE and total water extraction, which could be used as a proxy for root mass. This led us to conclude that the non inclusion of root mass in our TE calculation was unlikely to undermine much of the genetic differences in TE explained here.

In the current work, TE was highly correlated with total biomass ( $R^2 = 0.82$ , data not shown), which is similar to that found recently by Xin *et al.* (2009), but different from

Hammer *et al.* (1997). Also, the absence of relationship between TE and total water use, tested here in a large and representative set of germplasm, undermines previous speculation that TE and water use could be negatively related (Blum 2005) and clearly shows that it is possible to find germplasm capable of exhibiting both high water extraction and high TE, as previously shown (Peng and Krieg 1992). Indeed, both high water extraction and high TE were exhibited by variants of the Durra race in this study. When comparing races, it appeared that both Caudatum and Durra had overall higher values of TE than other landraces. Therefore, large genotypic variation for TE, especially in Caudatum and Durra races, could be exploited by sorghum breeding programs. The reasons for the superiority of the Caudatum and Durra races is unknown but we speculate that some could be in the environmental conditions in which these landraces have evolved. It has been recently shown that certain sorghum genotypes restrict transpiration at high vapor pressure deficit (Ghoolipoor *et al.*, 2010), which could lead to water saving and/or differences in transpiration efficiency.

#### Large differences in water extraction capacity

Little work has been done to assess water extraction *per se*, except for a detailed study on two genotypes by Robertson *et al.* (1993). Indeed, most studies in different crops so far have relied on assessing rooting characteristics and not on the function of root systems (Siddique *et al.* 1990; Sanguinetti *et al.* 1998; Kashiwagi *et al.* 2005). Therefore, our study may be the first to report a large range in variation for water extraction across an extensive germplasm set. Because of the lack of a strong relationship between water use under WW and DS conditions, water extraction differences were not related to constitutive traits but rather to differences in a response to stress. The low coefficient of variation (6%) for total water extraction measurements clearly indicates the value of the lysimetric system for assessing water extraction. In addition, it resolves previous complications related to identifying drought-adaptive rooting traits (Price *et al.* 2002).

These water extraction differences could relate to either a deeper rooting (Singh *et al.*, 2010), or to differences in the effective depth of water extraction. We estimate that each cylinder contains a total of 27 L of water (about 23% w/w is a typical value for this

Alfisol), from which about 70% can be extracted for transpiration (unpublished data). Therefore, approximately 19 L of water would then be available for extraction from the soil profile of the lysimeters, assuming the root length density was sufficient to do so. Hence, the sorghum genotypes extracting the most water from the profile, i.e. about 13 L after deducting 2L of water that were added in the course of the experiment, had roots sufficient to extract all possible water from about 70% of the soil profile. By contrast, genotypes extracting the least water would have attained full extraction to only about 50% of the soil profile. For the sake of representing these water extraction differences in terms of rooting depth, a 3 L plant<sup>-1</sup> difference represents all the water available in a 40 cm depth of the lysimeter soil profile.

Our method of assessing the pattern of water extraction during the whole life cycle is an innovation that adds precision and high throughput to existing field-based equipments (e.g. TDR, neutron probes). Rather than relying on static and destructive measurements of rooting characteristics, lysimeters sown with individual plants enabled water use to be evaluated under controlled water regimes in a dynamic manner. It revealed a large range of variation in both pre- and post-anthesis water use, and that these two parameters were highly and negatively correlated. Although both pre- and post-anthesis water use were related to flowering time, Figure 4 also clearly shows that for a similar level of pre-anthesis water use, there was still variation in post-anthesis water use, indicating that genotypes varied in their capacity to extract water during the grain filling period. More work is needed to assess how post-anthesis water use affects yield in genotypes with similar flowering time and pre-anthesis water use, as previously considered (Hammer 2006). The profile of water extraction also provided insight into when sorghum plants are getting stressed, i.e. not until about 4 weeks after the last irrigation. Although this type of information has been acquired from the field (Ritchie, 1981; Steiner, 1983, Turner et al. 1986), it is possible to acquire at a much larger scale and with more precision using the lysimetric system, enabling detailed studies of the relationships between patterns of water use and grain yield.

Besides HI, yield relates closely to TE and third only to water use



Yield variations under DS had no relation with the yield potential (Fig. 1). With a crop density of 11 plant m<sup>-2</sup> in the lysimetric system, the mean trial grain yield under terminal stress corresponded to 2.1 t ha<sup>-1</sup>, which was in the range of yield previously observed in the field with post-rainy sorghum materials of 1.2 – 2.1 t ha<sup>-1</sup> under similar weather conditions (Mahalakshmi and Bidinger 2002). Yields under fully irrigated conditions corresponded to 4.2 t ha<sup>-1</sup>, indicating that the stress imposed in the cylinders was within the desired range.

Among the components of Passioura's equation, HI explained the largest portion of the genetic variation in yield. This would be expected from germplasm with such a large range of variation for HI under both water regimes, and also from the fact that yield and HI have terms in common. Nevertheless, Figure 6 also showed that yield variation remained unexplained by HI. Our multi-linear approach, using a similar methodology to that in previous work (Bidinger *et al.* 1987; Vadez *et al.* 2007b), shows that the effect of HI on the yield variation needs to be removed before the strong effect of TE on the substantial residual variations can be highlighted. This reveals the important impact of TE on yield differences under terminal stress conditions in sorghum, as previously suggested (Hammer *et al.* 1997; Sinclair *et al.* 2005; Xin *et al.* 2009).

Interestingly, under well-watered conditions, the harvest index had much less of an influence on yield than under terminal stress ( $R^2 = 0.57$  vs  $R^2 = 0.88$ ). By contrast, both TE and water use (WU) had a significant effect on yield ( $R^2 = 0.34$  for TE;  $R^2 = 0.16$  for WU) when water was not limiting. In that case, we also computed the residual yield unexplained by HI. The correlation between these residuals and TE was also highly significant ( $R^2 = 0.29$ ,  $P < 0.01$ ), although it did not improve the direct relationship that was previously found between TE and yield. By contrast, the residuals were highly correlated to the water used ( $R^2 = 0.69$ ), with a significant improvement compared to the relationship previously drawn between WU and yield. These data then support the importance of maximizing water use when water is non-limiting, as previously suggested (Blum 2005).

These data highlight the potential of the lysimetric system to accurately assess the various components of Passioura's equation, allowing the respective weighting of their

importance under different watering regimes. Work is now in progress to test a number of water deficit conditions to assess the possible change in the bearing of each component. In addition, other traits also had a significant bearing on yield under terminal drought. In particular, extracting more water during the 59-78 DAS time interval, which corresponded to flowering and early grain filling, had a significant and positive effect on yield. This was despite the highly variable germplasm used in this work (germplasm with differences in tillering and harvest index). These data suggest that water uptake during that period may be critical for successful seed setting and grain filling of the crop, as previously shown and suggested (Ratnakumar *et al.* 2009; Vadez *et al.* 2007a; Zaman-Allah *et al.*, 2011), and also found in pearl millet (Vadez *et al.* 2009).

Summarising, the lysimetric system was suitable for a) generating yield data that approximate field conditions, and b) assessing the value of the various components of the Passioura equation on yield. The use of either step-wise regression or multi-linear regression analysis was needed to remove components of greater influence in order to highlight the significant influence of other traits.

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## References

- Balota M, Payne WA, Rooney W, Rosenow D (2008) Gas exchange and transpiration ratio in sorghum. *Crop Science* **48**, 2361-2371.
- Bhan S, Singh HG, Singh A (1973) Note on root development as an index of drought resistance in sorghum (*Sorghum bicolor* L. Moench). *Indian Journal of Agricultural Sciences* **43**, 828-830.

- 538 Bidinger FR, Mahalakshmi V, Rao GDP (1987) Assessment of drought resistance in  
539 pearl millet (*Pennisetum americanum* (L.) Lecke). II: Estimation of genotype  
540 response to stress. *Australian Journal of Agricultural Research* **38**, 37-48.
- 541 Blum A (2005) Drought resistance, water use efficiency, and yield potential – Are they  
542 compatible, dissonant, or mutually exclusive. *Australian Journal of Agricultural*  
543 *Research* **56**, 1159-1168.
- 544 Blum A, Jordan WR, Arkin GF (1977) Sorghum Root Morphogenesis and Growth. II.  
545 Manifestation of Heterosis. *Crop Science* **17**, 153-157.
- 546 Crasta OR, Xu WW, Rosenow DT, Mullet J, Nguyen HT (1999) Mapping of post-  
547 flowering drought resistance traits in grain sorghum: association between QTLs  
548 influencing premature senescence and maturity. *Molecular and General Genetics*  
549 **262**, 579–588.
- 550 Donatelli M, Hammer GL, Vanderlip RL (1992) Genotype and water limitation effects on  
551 phenology, growth, and transpiration efficiency in grain sorghum. *Crop Science* **32**,  
552 781–786.
- 553 Hammer GL, Farquhar GD, Broad IJ (1997) On the extent of genetic variation for  
554 transpiration efficiency in sorghum. *Australian Journal of Agricultural Research*  
555 **48**, 649–655.
- 556 Hammer GL (2006) Pathways to prosperity: Breaking the yield barrier in sorghum. In:  
557 A.K. Borrell and D.R. Jordan, *5th Australian Sorghum Conference*, Gold Coast,  
558 Qld, Australia, (1-19).
- 559 Harris K, Subudhi PK, Borrell AK, Jordan DB, Rosenow D, Nguyen H, Klein P, Klein R,  
560 Mullet J (2007) Sorghum stay-green QTL individually reduce post-flowering  
561 drought-induced leaf senescence. *Journal of Experimental Botany* **58**, 327-338.
- 562 Hubick KT, Farquhar GD, Shorter R (1986) Correlation between water-use efficiency  
563 and carbon isotope discrimination in diverse peanut (*Arachis*) germplasm.  
564 *Australian Journal of Plant Physiology* **13**, 803-816.
- 565 Jordan WR, Miller FR, Morris DE (1979) Genetic Variation in Root and Shoot Growth of  
566 Sorghum in Hydroponics. *Crop Science* **19**, 468-472.

- 567 Kashiwagi J, Krishnamurthy L, Upadhyaya HD, Krishna H, Chandra S, Vadez V,  
568 Serraj R (2005) Genetic variability of drought-avoidance root traits in the mini-core  
569 germplasm collection of chickpea (*Cicer arietinum* L.). *Euphytica* **146**, 213-222.
- 570 Kirkegaard JA, Lilley JM, Howe GN, Graham JM (2007) Impact of subsoil water use on  
571 wheat yield. *Australian Journal of Agricultural Research* doi:10.1071/AR06285
- 572 Kholová J, Hash CT, Kočvá M, Vadez V (2010a) Constitutive water conserving  
573 mechanisms are correlated with the terminal drought tolerance of pearl millet  
574 (*Pennisetum americanum* L.). *Journal of Experimental Botany* **61**, 369–377.
- 575 Kholová J, Hash CT, Kumar LK, Yadav RS, Kočvá M, Vadez V (2010b) Terminal  
576 drought tolerant pearl millet [*Pennisetum glaucum* (L.) R. Br.] have high leaf ABA  
577 and limit transpiration at high vapor pressure deficit *Journal of Experimental*  
578 *Botany* **61**, 1431-1440.
- 579 Kirkegaard JA, Lilley JM, Howe GN, Graham JM (2007) Impact of subsoil water use on  
580 wheat yield. *Australian Journal of Agricultural Research* **58**, 303-315
- 581 Krishnamurthy L, Vadez V, Devi MJ, Serraj R, Nigam SN, Sheshshayee MS, Chandra S,  
582 Aruna R (2007) Variation in transpiration efficiency and its related traits in a  
583 groundnut (*Arachis hypogaea* L.) mapping population. *Field Crops Research* **103**,  
584 189-197.
- 585 Mahalakshmi V, Bidinger FR (2002) Evaluation of staygreen sorghum germplasm lines  
586 at ICRISAT. *Crop Science* **42**, 965-974.
- 587 Manschadi AM, Christopher JT, Peter deVoil P, Hammer GL (2006) The role of root  
588 architectural traits in adaptation of wheat to water-limited environments. *Functional*  
589 *Plant Biology* **33**, 823-837.
- 590 Mayaki WC, Stone LR, Teare ID (1976) Irrigated and Nonirrigated Soybean, Corn, and  
591 Grain Sorghum Root Systems. *Agronomy Journal* **68**, 532-534.
- 592 Nageswara Rao, RC, Talwar HS, Wright GC (2001) Rapid assessment of specific leaf  
593 area and leaf nitrogen in peanut (*Arachis hypogaea* L.) using a chlorophyll meter.  
594 *Journal of Agronomy and Crop Science* **186**, 175-182.

- 595 Passioura JB (1977) Grain yield, harvest index and water use of wheat. *Journal of*  
596 *Australian Institute of Agriculture Science* **43**, 117—121.
- 597 Peng S, Krieg DR (1992) Gas exchange traits and their relationship to water use  
598 efficiency. *Crop Science* **32**, 386–391.
- 599 Price AH, Townend J, Jones MP, Audebert A, Courtois B (2002) Mapping QTLs  
600 associated with drought avoidance in upland rice grown in the Philippines and West  
601 Africa. *Plant Molecular Biology* **48**, 683—695.
- 602 Ratnakumar P, Vadez V, Nigam SN, Krishnamurthy L (2009) Assessment of  
603 transpiration efficiency in peanut (*Arachis hypogaea* L.) under drought by  
604 lysimetric system. *Plant Biology* **11**, 124-130.
- 605 Ritchie JT (1981) Water dynamics in the soil-plant-atmosphere system. *Plant and Soil*  
606 **58**, 81-96.
- 607 Robertson MJ, Fukai S, Ludlow MM, Hammer GL (1993) Water extraction by grain  
608 sorghum in a subhumid environment . 1. Analysis of the water extraction pattern.  
609 *Field Crops Research* **33**, 81-97.
- 610 Salih AA, Ali IA, Lux A, Luxova M, Cohen Y, Sugimoto Y, Inanga S (1999) Rooting,  
611 water uptake, and xylem structure adaptation to drought of two sorghum cultivars.  
612 *Crop Science* **39**, 168-173.
- 613 Sanguineti MC, Giuliani MM, Govi G, Tuberosa R, Landi P (1998) Root and shoot traits  
614 of maize inbred lines grown in the field and in hydroponic culture and their  
615 relationships with root lodging. *Maydica* **43**, 211-216.
- 616 Serraj R, Hash CT, Buhariwalla HK, Bidinger FR, Folkertsma RT, Chandra S, Gaur PM,  
617 Kashiwagi J, Nigam SN, Rupakula A, Crouch JH (2005). Marker-assisted breeding  
618 for crop drought tolerance at ICRISAT: Achievements and prospects. Pages 217-  
619 238 in In the Wake of the Double Helix: From the Green Revolution to the Gene  
620 Revolution (Tuberosa R, Phillips RL and Gale MD, eds.). Bologna , Italy : Avenue  
621 media.
- 622 Siddique KHM, Belford RK, Tennant D (1990) Root:shoot of. old and modern, tall and  
623 semi-dwarf wheat in Mediterranean-. environment. *Plant and Soil* **121**, 89-98.

- 624 Sinclair TR, Muchow CR (2001) System analysis of plant traits to increase grain yield on  
625 limited water supplies. *Agronomy Journal* **93**, 263—270.
- 626 Sinclair TR, Hammer GL, van Oosterom EJ (2005) Potential yield and water-use  
627 efficiency benefits in sorghum from limited maximum transpiration rate. *Functional*  
628 *Plant Biology* **32**, 945–952.
- 629 Singh V, van Oosterom EJ, Jordan DR, Messina CD, Cooper M, Hammer GL (2010)  
630 Morphological and architectural development of root systems in sorghum and  
631 maize. *Plant and Soil* **333**, 287-299.
- 632 Steiner JL (1983) Dryland grain sorghum water use, light interception, and growth  
633 response to planting geometry. *Agronomy Journal* **78**, 720-726.
- 634 Turner NC, Hearn AB, Begg JE and Constable GA (1986) Cotton (*Gossypium hirsutum*  
635 L.): Physiological and morphological responses to water deficits and their  
636 relationship to yield *Field Crops Research* **14**, 153-170.
- 637 Udayakumar M, Sheshshayee MS, Nataraj KN, Bindu Madhava H, Devendra R, Aftab  
638 Hussain IF, Prasad TG (1998) Why has breeding for water-use efficiency not been  
639 successful? An analysis and alternate approach to exploit this trait for crop  
640 improvement. *Current Science* **74**, 994-1000.
- 641 Upadhyaya HD, Pundir RPS, Dwiwedi SL, Gowda CLL, Reddy VG, Singh S (2009)  
642 Developing a mini core collection of sorghum for diversified utilization of  
643 germplasm. *Crop Science* **49**, 1769-1780.
- 644 Vadez V, Krishnamurthy L, Kashiwagi JW, Kholova J, Devi JM, Sharma KK,  
645 Bhatnagar-Mathur P, Hoisington DA, Hash CT, Bidinger FR, Keatinge JDH  
646 (2007a) Exploiting the functionality of root systems for dry, saline, and nutrient  
647 deficient environments in a changing climate. *J. SAT Agric Res* Vol **4** (Special  
648 Symposium edition) <http://www.icrisat.org/journal/specialproject.htm>
- 649 Vadez V, Krishnamurthy L, Gaur PM, Upadhyaya HD, Hoisington DA, Varshney RK,  
650 Turner NC, Siddique KHM (2007b) Large variation in salinity tolerance is  
651 explained by differences in the sensitivity of reproductive stages in chickpea. *Field*  
652 *Crop Research* **104**, 123–129.

- 653 Vadez V, Rao S, Kholova J, Krishnamurthy L, Kashiwagi J, Ratnakumar P, Sharma KK,  
654 Bhatnagar-Mathur P, Basu PS (2008) Roots research for legume tolerance to  
655 drought: *Quo vadis? Journal of Food Legumes* **21** (2) 77-85.
- 656 Vadez V, Kholová J, Kakker A, Hash CT, Yadav R, Kočová M (2009) Pearl millet  
657 genotypes differing for a terminal drought tolerance QTL contrast for traits related  
658 to the control of leaf water loss. P170 in the proceedings from the Interdrought III  
659 conference, Oct 11-16, Shanghai Academy of Agriculture science, Shanghai, China.
- 660 Wright GC, Nageswara Rao RC, Farquhar GD (1994) Water-use efficiency and carbon  
661 isotope discrimination in peanut under water deficit conditions. *Crop Science* **34**,  
662 92-97.
- 663 Xin Z, Aiken R, Burke JJ (2009) Genetic diversity of transpiration efficiency in sorghum  
664 *Field Crops Research* **111**, 74-80.
- 665 Zaman-Allah M, Jenkinson D, Vadez V (2011). A conservative pattern of water use,  
666 rather than deep or profuse rooting, is critical for the terminal drought tolerance of  
667 chickpea. *Journal of Experimental Botany*. doi: 10.1093/jxb/err139  
668

**Table 1. Trial means, range of expected means, standard error of differences (SED) within treatment, and wald statistics and F-probability for genotype effect (G), treatment effect (T) and genotype-by-treatment (GxT) interaction related to time to flowering (d), grain dry mass (g plant<sup>-1</sup>), total dry mass (g plant<sup>-1</sup>), harvest index (HI), transpiration efficiency (TE, g kg<sup>-1</sup>) and panicle harvest index (PNHI, i.e. the ratio of the grain weight by the panicle weight)**

		50% FI		Grain yield		Total DW		Harvest index		TE		PNHI	
		WW	DS	WW	DS	WW	DS	WW	DS	WW	DS	WW	DS
Mean		57	56	41.97	20.59	126.94	67.05	0.33	0.27	4.38	4.82	0.75	0.70
Min		46	44	2.06	0.23	34.01	41.75	0.02	0.00	2.95	3.21	0.26	0.00
Max		67	72	82.83	36.76	193.29	88.47	0.53	0.45	5.59	6.09	0.89	0.89
$\sigma_g^2$		13.1	17.6	166.0	38.69	664.6	43.47	0.06173	0.00584	0.162	0.162	0.005493	0.00623
SE		1.9	2.3	28.2	6.53	106.3	8.89	0.00098	0.00095	0.054	0.038	0.00099	0.00118
SED		2.14	2.02	10.07	4.80	18.83	5.99	0.057	0.056	0.45	0.39	0.061	0.067
G	$\sigma_g^2$	15.0		41.6		113.2		0.00496		0.113		0.005677	
	SE	1.94		12.5		40.7		0.00080		0.032		0.000942	
T	Wald	14.6		325		731.6		56.5		78.4		33.1	
	Prob	0.001		0.001		0.001		0.001		0.01		0.001	
G x T	$\sigma_{gxT}^2$	0.65		61.1		236.7		0.001342		0.049		0.001065	
	SE	0.42		12.4		43.7		0.000433		0.032		0.000540	



**Table 2. Trial means, range of expected means, standard error of differences (SED) within treatment, and wald statistics and F-probability for genotype effect (G), treatment effect (T) and genotype-by-treatment (GxT) interaction related to total water use (g plant<sup>-1</sup>), pre-anthesis water use (g plant<sup>-1</sup>), post anthesis water use (g plant<sup>-1</sup>), and water used in the 45-59 DAS (g plant<sup>-1</sup> d<sup>-1</sup>), 59-78 DAS (g plant<sup>-1</sup> d<sup>-1</sup>) and 78-94 DAS (g plant<sup>-1</sup> d<sup>-1</sup>) periods**

Water use		Total		Pre-anthesis		Post-anthesis		45-59 DAS		59-78 DAS		78-94 DAS	
		WW	DS	WW	DS	WW	DS	WW	DS	WW	DS	WW	DS
Mean		28,856	13,908	9,383	8,956	20,727	6,452	358	344	401	180	368	112
Min		10,500	10,620	5,406	4,493	5,854	2,439	141	162	114	70	99	58
Max		42,320	15,240	15,493	13,628	35,018	10,917	556	668	598	279	630	229
$\sigma^2_g$		27,908,726	158,959	3,641,052	1,731,958	22,464,015	1,982,010	4,528	1,452	4,482	677	7,954	334
SE		4,141,939	50,275	535,809	285,982	3,435,089	301,436	759	969	838	260	1,164	76.7
SED		3,467	436	1,147	999	3,053	961	52	49	57	30	57	18
G	$\sigma^2_g$	1,124,515		2,183,060		1,452,707		2641		974		75	
	SE	1,456,941		328,263		1,238,507		600		375		430	
T	Wald	1,017		8.95		1,132		4.19		702		953	
	Prob	0.001		0.003		0.001		0.04		0.001		0.001	
G x T	$\sigma^2_{g \times T}$	12,814,903		365,709		10,488,093		247		3509		4187	
	SE	1,939,986		144,457		1,572,411		53.9		641		620	

**Table 3. Multi-linear regression between yield and several explanatory variables under DS conditions: harvest index, transpiration efficiency, total water extracted, post-anthesis water use, water used in the 45-59 DAS and 59-78 DAS periods, and days to flowering**

Factors	Coefficient	SE	t-value	P > t
Harvest index	67.6	0.77	87.73	0.000
Transpiration efficiency	4.28	0.14	29.59	0.000
Total water extracted	0.00163	0.00015	10.90	0.000
Post-anthesis water use	-0.00038	0.00017	-2.30	0.023
Water use between 45-59 DAS	0.00011	0.00015	1.81	Ns
Water use between 59-78 DAS	0.00045	0.00019	2.32	0.021
Days to 50% flowering	-0.161	0.053	-3.02	0.003
Constant	-34.30	3.92	-8.74	0.000

**Table 4. Multi-linear regression between the residual yield variations not explained by HI and several explanatory variables under DS conditions: transpiration efficiency, total water extracted, post-anthesis water use, water used in the 45-59 DAS and 59-78 DAS periods, and days to flowering**

Factors	Coefficient	SE	t-value	P > t
Transpiration efficiency	3.44	0.18	18.49	0.000
Total water extracted	0.00147	0.00019	7.77	0.000
Post-anthesis water use	-0.00044	0.00022	-2.02	0.046
Water use between 45-59 DAS	0.00029	0.00012	2.39	0.018
Water use between 59-78 DAS	0.00066	0.00025	2.64	0.009
Days to 50% flowering	-0.105	0.069	-1.53	ns
Constant	-32.12	5.11	-6.28	0.000

### Figure Captions

**Fig. 1.** Relationship between grain yield under terminal water stress ( $\text{g plant}^{-1}$ ) and grain yield under well-watered conditions ( $\text{g plant}^{-1}$ ) (a), and relationship between harvest index (HI) under terminal water stress and HI under well-watered conditions (b) in 152 germplasm entries. Data are the mean of 3 replicated lysimeter-grown plants per genotype.

**Fig. 2.** Total water extracted from the lysimeter soil profile ( $\text{g plant}^{-1}$ ) under terminal water stress conditions in the different sorghum races. Data are the mean of the average transpiration values within each race (Bicolor,  $n=17$ ; Caudatum,  $n=31$ ; Caudatum-Bicolor (C-B),  $n=14$ ; Durra,  $n=18$ ; Durra-Caudatum (D-C),  $n=13$ ; Guinea,  $n=14$ ; Guinea-Caudatum (G-C),  $n=24$ ; Kafir,  $n=6$ ).

**Fig. 3.** Profile of transpiration ( $\text{g plant}^{-1}$ ) as a function of time after sowing. Last irrigation was applied at 30 DAS in plants exposed to terminal water stress (DS, open symbols) and well-watered conditions (WW, closed symbols). Data are the mean ( $\pm$  SE) of the average transpiration values for 152 germplasm entries. For the DS and WW plants, five and six lysimeter weighings, respectively, were done, giving four and five transpiration intervals.

**Fig. 4.** Relationship between the pre-anthesis water use ( $\text{g plant}^{-1}$ ) and the post-anthesis water use ( $\text{g plant}^{-1}$ ) in 152 germplasm entries. Data are the mean of 3 replicated lysimeter-grown plants per genotype.

**Fig. 5.** Transpiration efficiency ( $\text{g kg}^{-1}$  water transpired) under terminal water stress conditions in the different sorghum races. Data are the mean of the average transpiration values within each race (Bicolor,  $n=17$ ; Caudatum,  $n=31$ ; Caudatum-Bicolor (C-B),  $n=14$ ; Durra,  $n=18$ ; Durra-Caudatum (D-C),  $n=13$ ; Guinea,  $n=14$ ; Guinea-Caudatum (G-C),  $n=24$ ; Kafir,  $n=6$ ).

**Fig. 6.** Relationship between (a) seed yield under terminal water stress and the harvest index (HI), (b) relationship between the residual yield variations unexplained by HI and transpiration efficiency (TE), and (c) relationship between the residual yield variations unexplained by either HI or TE and the total water extracted from the soil profile in 152 germplasm entries. Data are the mean of 3 replicated lysimeter-grown plants per genotype.

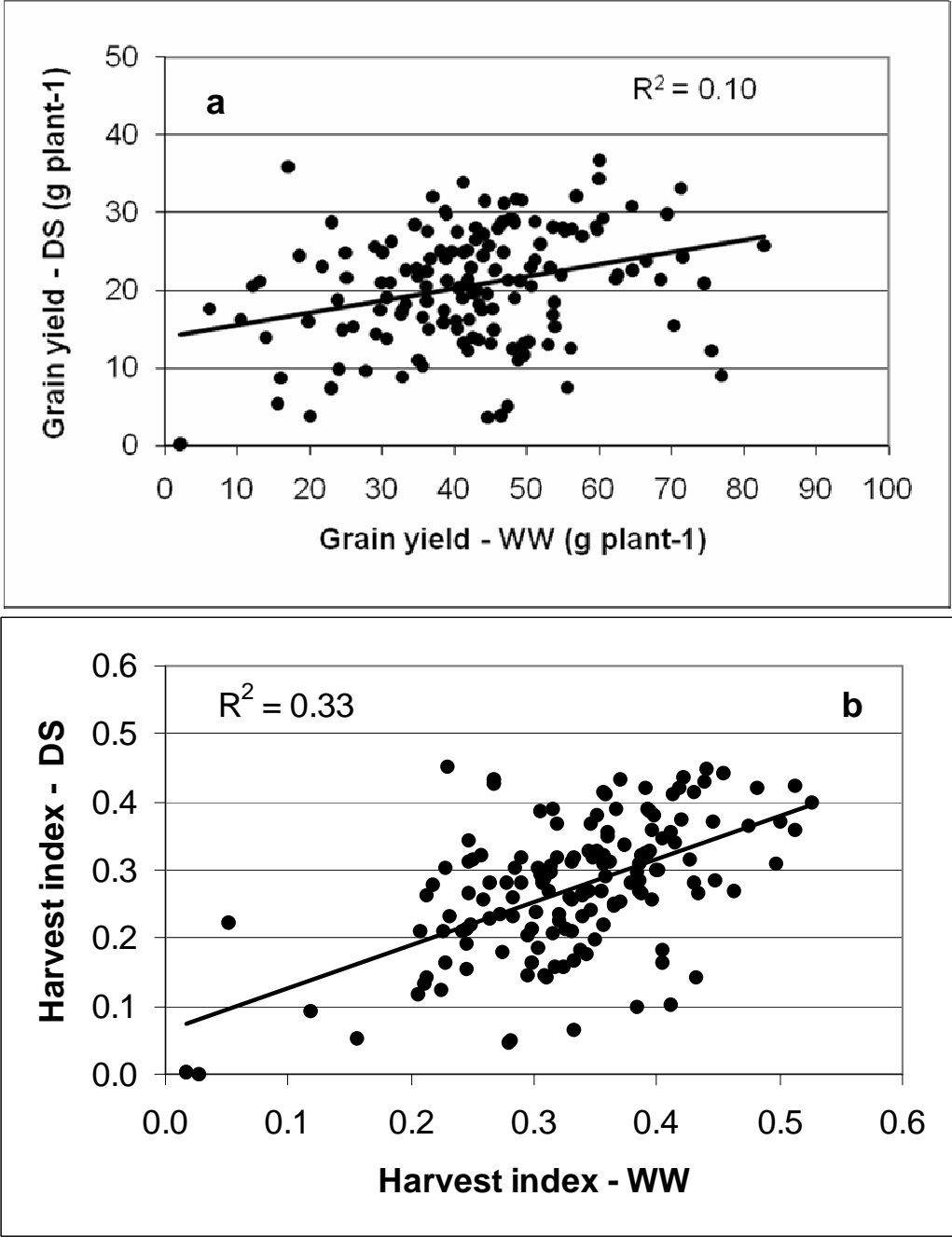


Figure 1

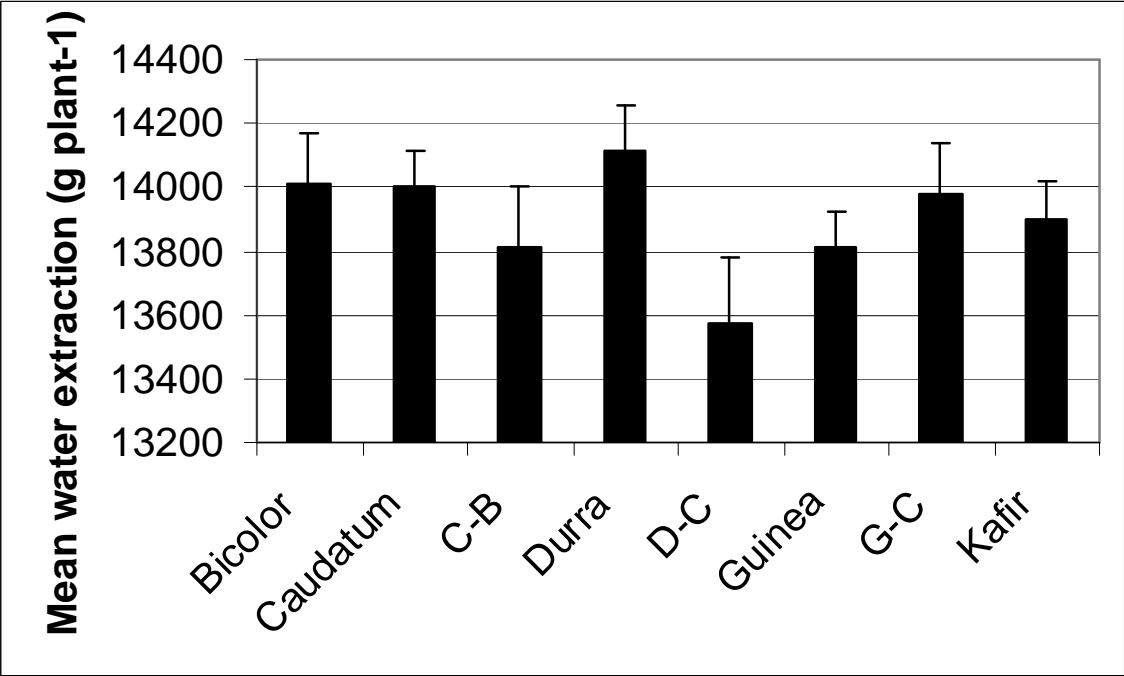


Figure 2

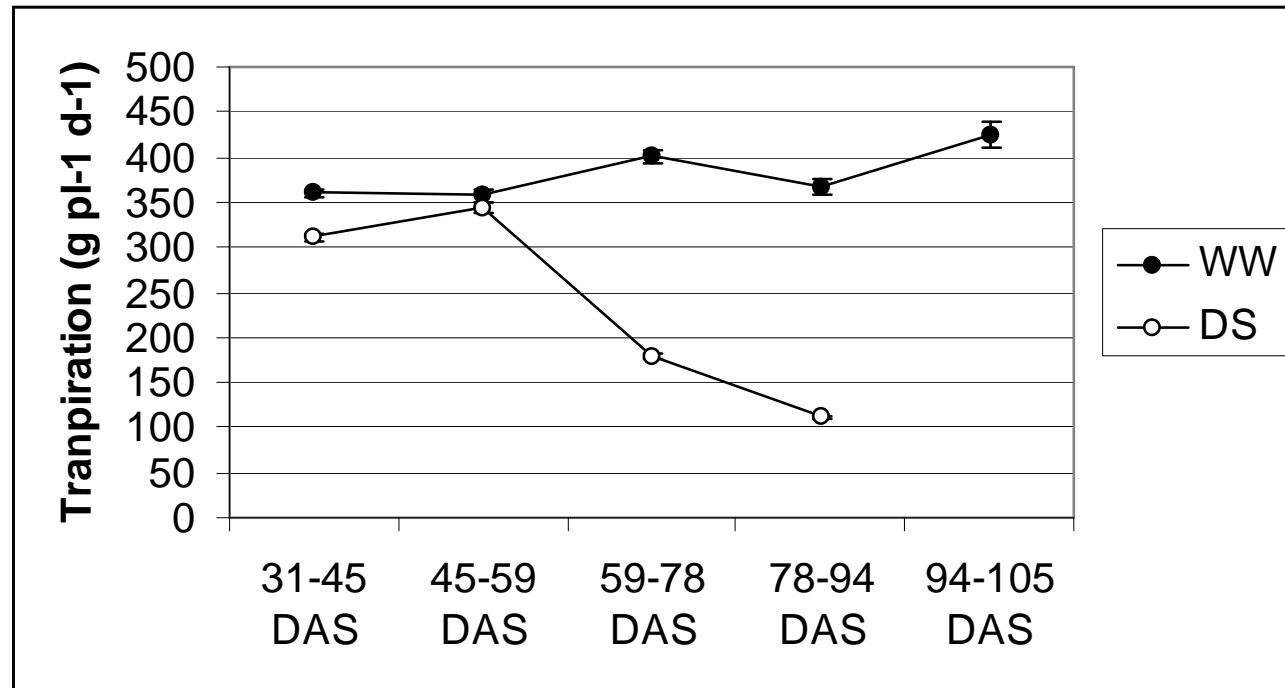


Figure 3

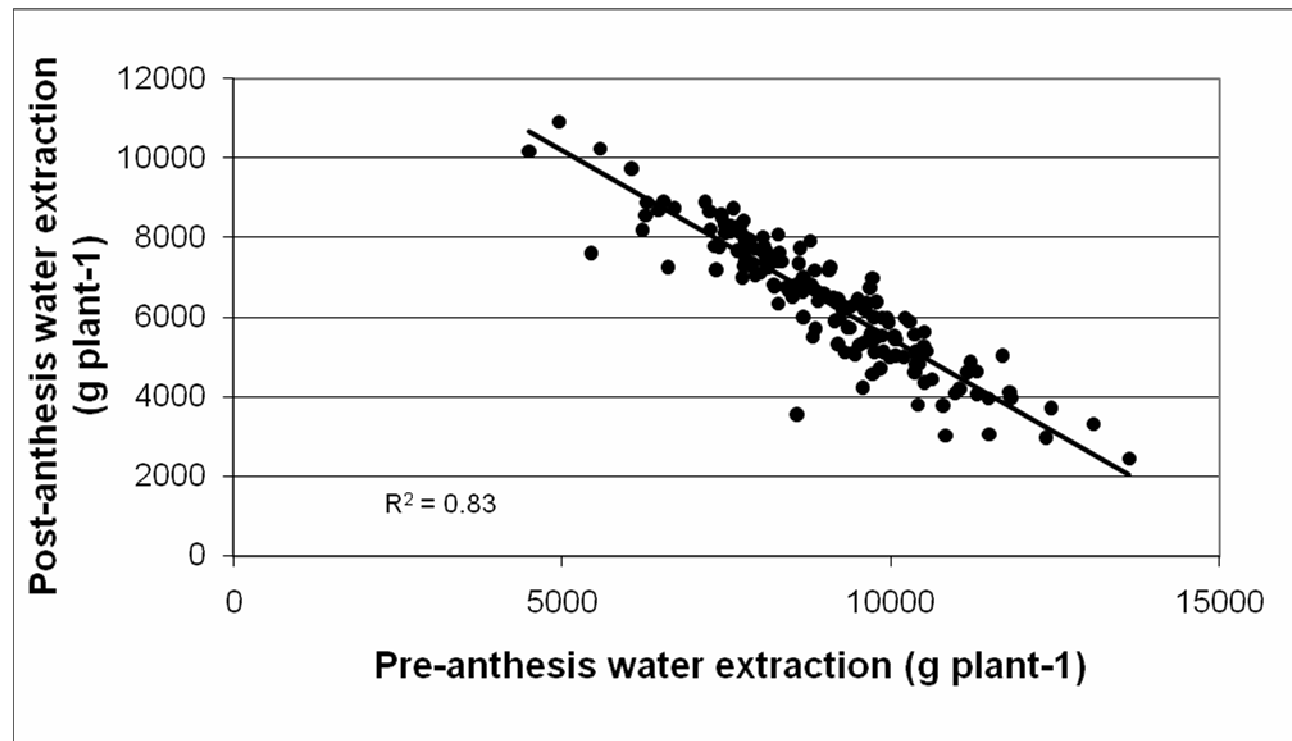


Figure 4



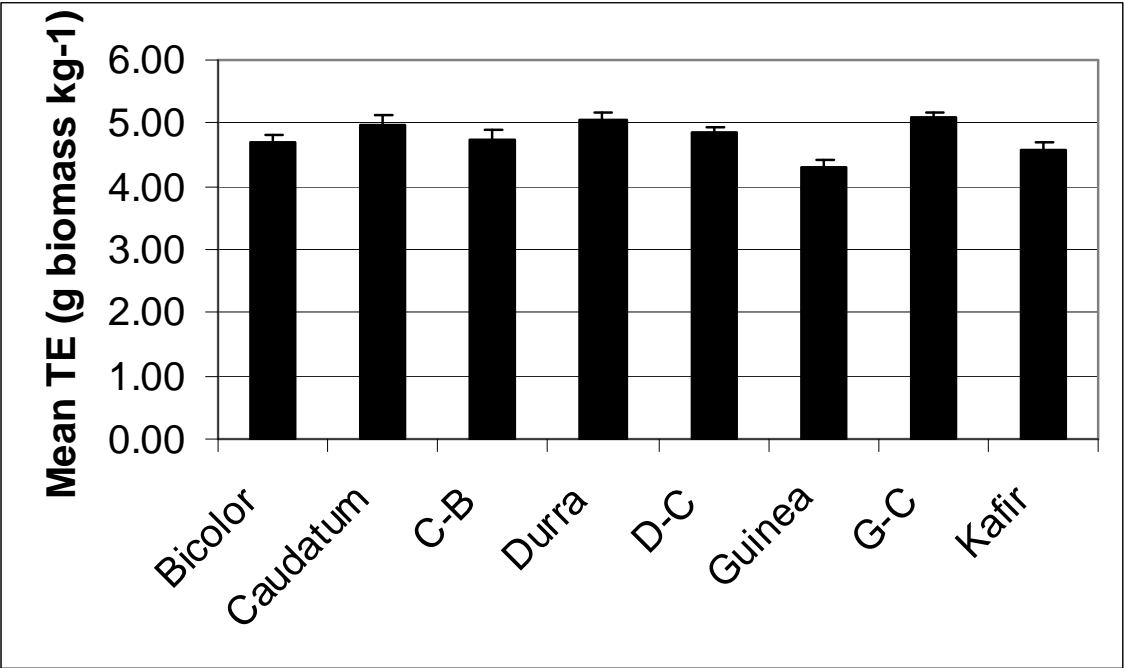


Figure 5

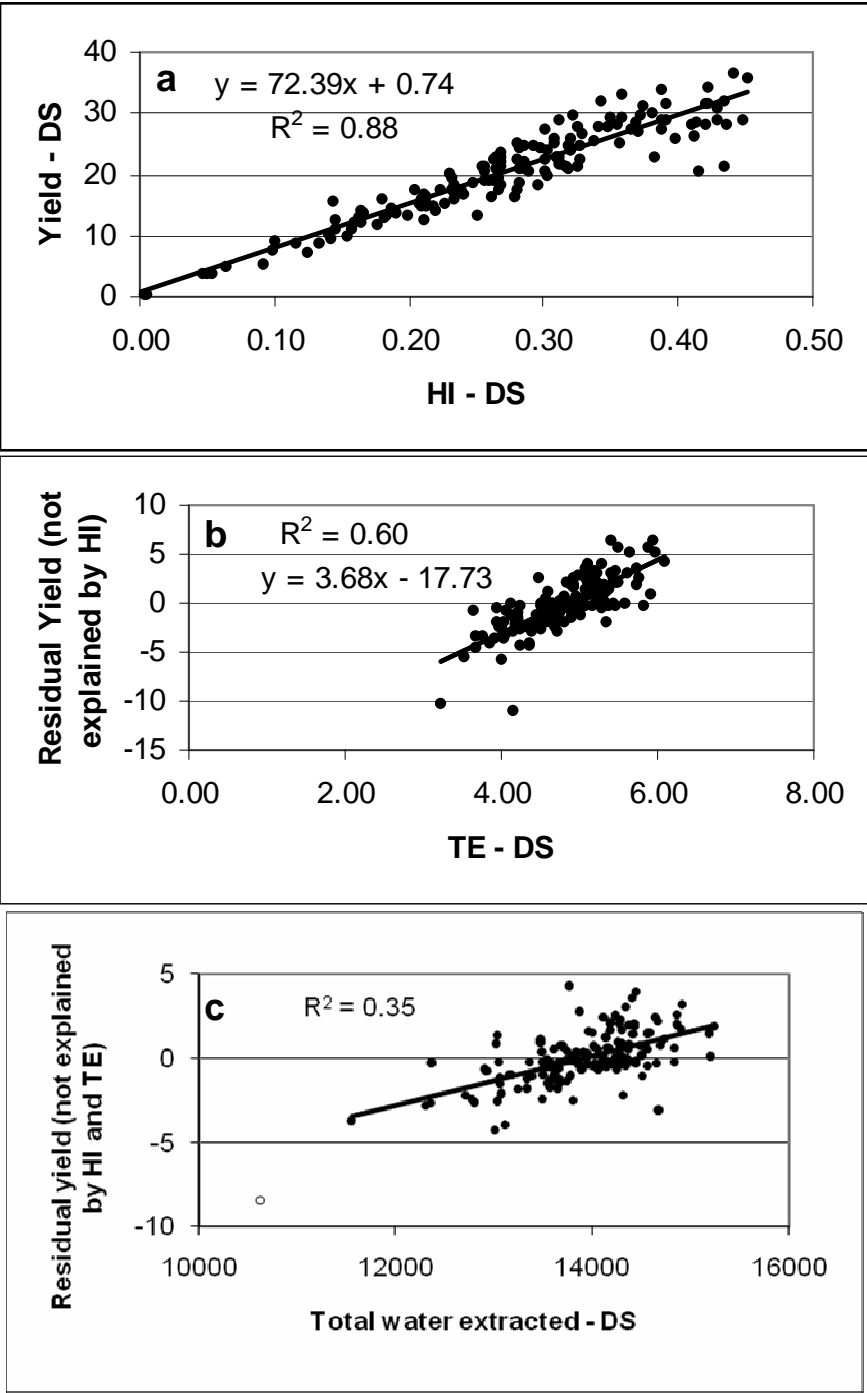


Figure 6